

# How does clinical data fit into two leukopoiesis models?

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# Motivation

# Blood cells production and regulation

## Motivation

- Haematopoiesis
- Differentiation stages
- Blood pathologies

## Two leukopoiesis models

## Solution methods

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## Concluding remarks

**Haematopoietic pluripotent stem cells** (HSCs) in bone marrow give birth to the three blood cell types.

**Growth factors** or **Colony Stimulating Factors** (CSF) – specific proteins that stimulate the production and maturation of each blood cell type.

**Blast cells** – blood cells that have not yet matured.

Blood cell type	Function	Growth factors
Erythrocyte	Transport oxygen to tissues	Erythropoietin
Leukocyte	Fight infections	G-CSF, M-CSF, GM-CSF, Interleukins
Thrombocyte	Control bleeding	Thrombopoietin

**Leukopoiesis** – process of production and regulation of white blood cells (T- and B-lymphocytes, NK cells, monocytes, granulocytes, eosinophils, and basophils)

# Differentiation stages in haematopoiesis

## Motivation

- Haematopoiesis
- Differentiation stages
- Blood pathologies

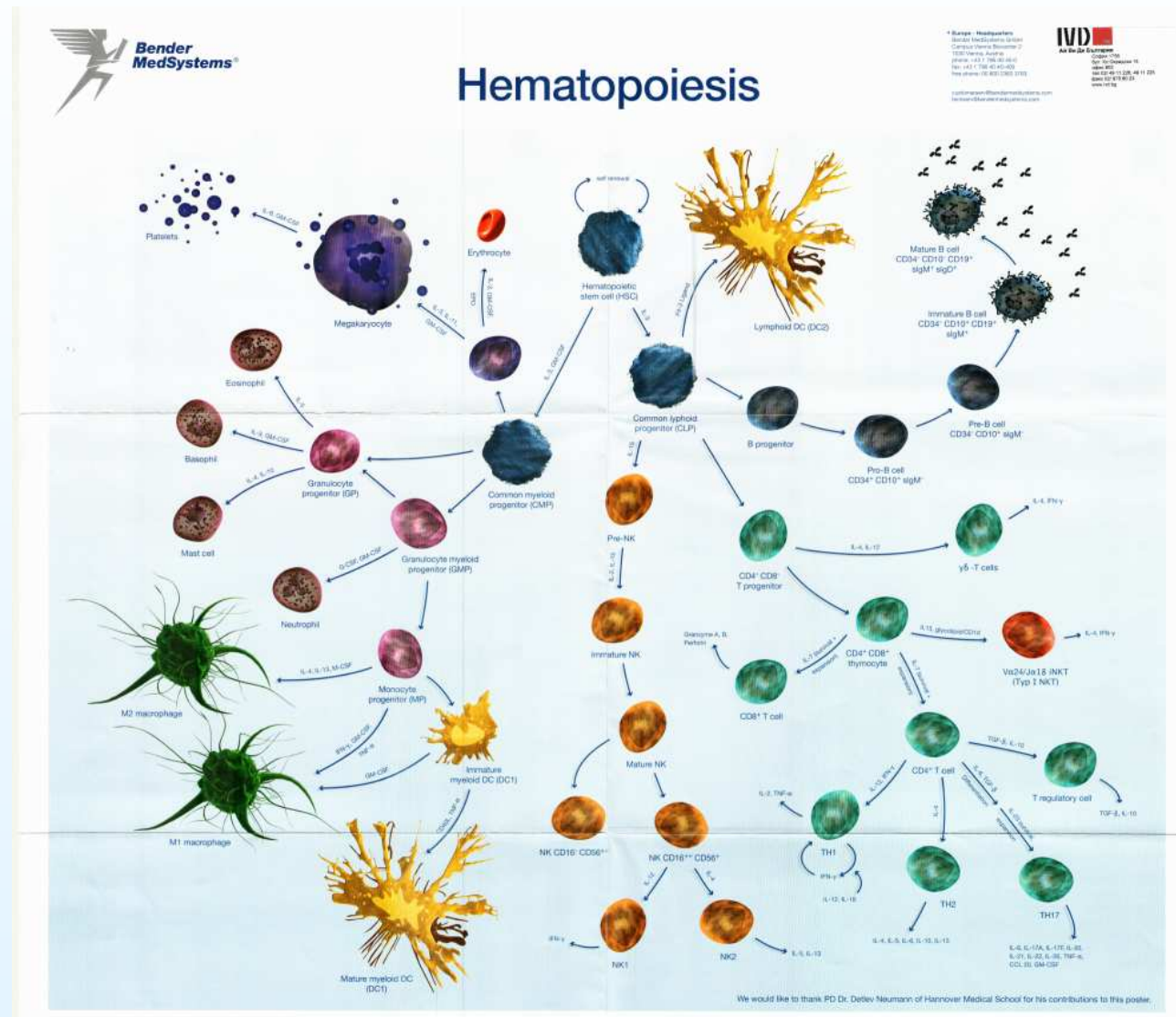
## Two leukopoiesis models

## Solution methods

## Clinical data

## Numerical tests

## Concluding remarks



<http://www.bendermedsystems.com/>

# Blood pathologies

Various **hematological diseases** (including leukemia) are characterized by **abnormal production** of particular blood cells (matured or blast).

Main stages in the therapy of blood diseases:

**TBI:** Total body irradiation (TBI) and chemotherapy – kill the "tumour" cells, but also the healthy ones.

**BMT:** Bone marrow transplantation (BMT) – stem cells of a donor (collected under special conditions) are put in the peripheral blood, from where they have to:

- 1) find their way to the stem cell niche in the bone marrow; and afterwards
- 2) selfrenew and differentiate to regenerate the patient's blood system.

**Current stage:** Compare clinical data and computer simulations with two leukopoiesis models.

## Motivation

- Haematopoiesis
- Differentiation stages
- **Blood pathologies**

## Two leukopoiesis models

## Solution methods

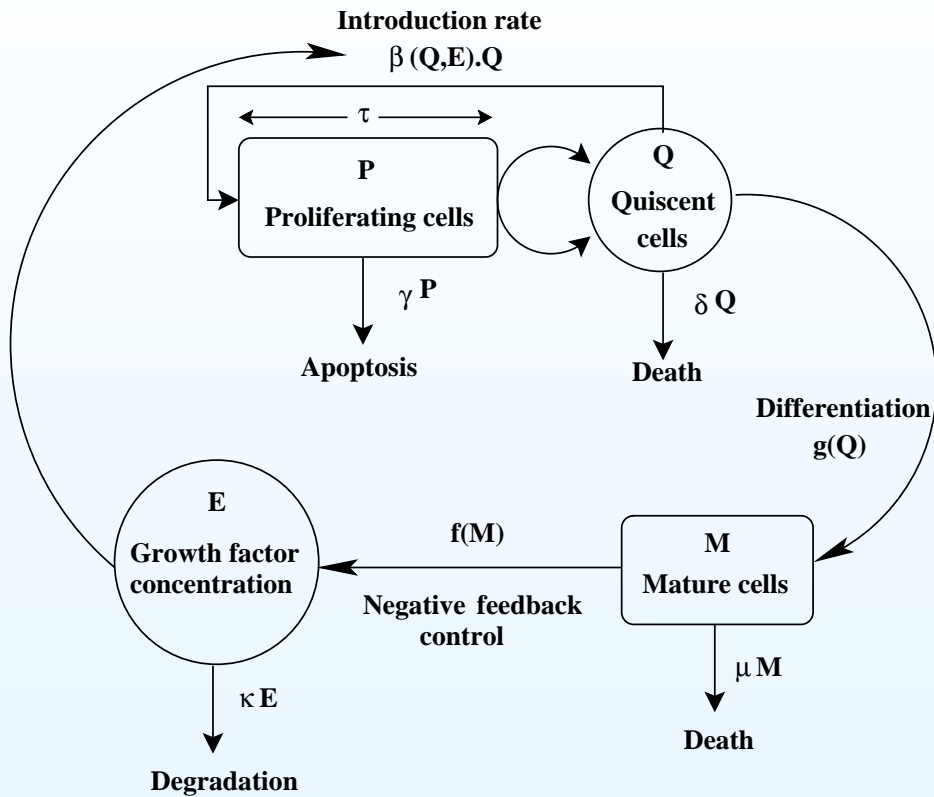
## Clinical data

## Numerical tests

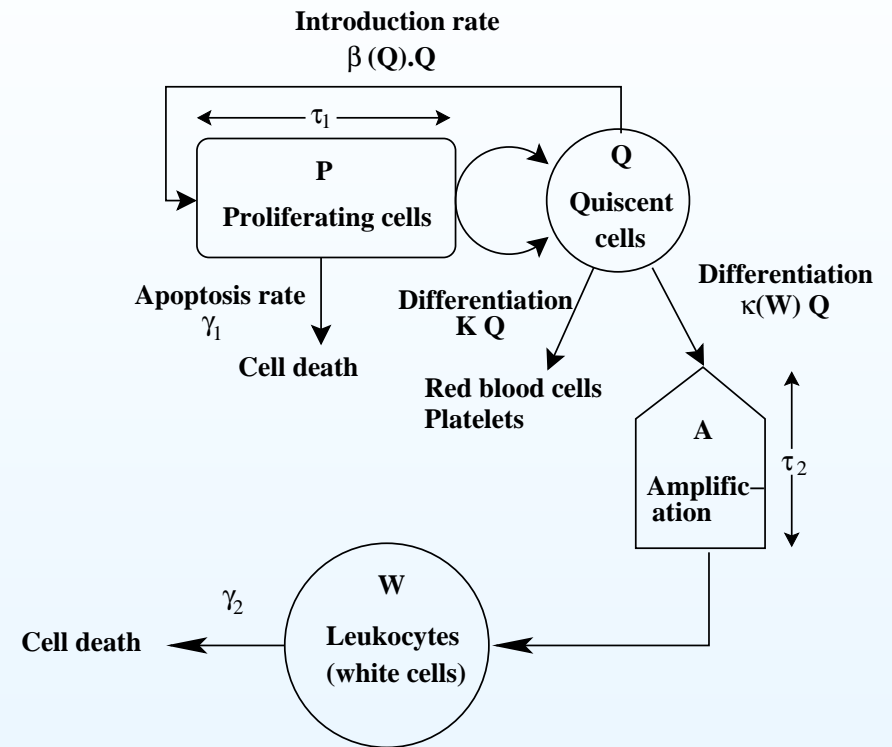
## Concluding remarks

# Two leukopoiesis models

# Involved data



Growth factors model (GFM)



Leukopoiesis model (LM)

[GFM] M. Adimy, F. Crauste, S. Ruan, *Modelling Hematopoiesis Mediated by Growth Factors with Applications to Periodic Hematological Diseases*, *Bulletin of Mathematical Biology*, 68 (8), (2006), 2321–2351.

[LM] M. Adimy, F. Crauste, S. Ruan, *Periodic oscillations in leukopoiesis models with two delays*, *Journal of Theoretical Biology* 242, (2006), 288–299.



# Description of parameters and functions

Stem cells –  $P$  in proliferating phase,  $Q$  in quiescent phase  
Growth factor –  $E$ , Mature cells –  $M$ , Leukocytes –  $W$

Proliferating phase duration –  $\tau, \tau_1$

Amplification phase duration –  $\tau_2$

Amplification parameter –  $A = \alpha 2^i$ , with

$\alpha \in (0, 1)$  – survival rate,  $i$  – number of generations

Apoptosis rate –  $\gamma, \gamma_1$

Death rate –  $\kappa$  (for  $E$ ),  $\mu$  (for  $M$ ),  $\gamma_2$  (for  $W$ ),  $\delta$  (for  $Q$ )

Introduction rate –  $\beta(Q, E), \beta(Q)$

Differentiation –  $g(Q), K, k(W)$

Motivation

Two leukopoiesis models

- Involved data
- **Parameters and functions**
- GFM system of DDEs
- LM system of DDEs

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# GFM system of DDEs

Motivation

Two leukopoiesis models

- Involved data
- Parameters and functions
- GFM system of DDEs
- LM system of DDEs

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$$(GFM) \left\{ \begin{array}{l} \frac{dQ}{dt} = -\delta Q(t) - g(Q(t)) - \beta(Q(t), E(t)) Q(t) \\ \quad \quad \quad + 2e^{-\gamma\tau} \beta(Q(t-\tau), E(t-\tau)) Q(t-\tau) \\ \frac{dM}{dt} = -\mu M(t) + g(Q(t)) \\ \frac{dE}{dt} = -kE(t) + f(M(t)) \end{array} \right.$$

$$Q(t) = Q_0(t), M(t) = M_0(t), E(t) = E_0(t), t \in [-\tau, 0]$$

Delay  $\tau$  corresponds to the cell cycle duration.

$$Q(t) \geq 0, M(t) \geq 0, E(t) \geq 0, k > 0, \mu > 0$$

Existence of nontrivial positive steady-state is ensured by:

$$0 < \delta + g'(0) < \beta\left(0, \frac{f(0)}{k}\right) \text{ and}$$

$$0 \leq \tau < \tau_{max} := \frac{1}{\gamma} \ln \left( \frac{2\beta\left(0, \frac{f(0)}{k}\right)}{\delta + g'(0) + \beta\left(0, \frac{f(0)}{k}\right)} \right)$$

# LM system of DDEs

Motivation

Two leukopoiesis models

- Involved data
- Parameters and functions
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- LM system of DDEs

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$$(LM) \begin{cases} \frac{dQ}{dt} = -[K + k(W(t)) + \beta(Q(t))]Q(t) \\ \quad + 2e^{-\gamma_1\tau_1}\beta(Q(t-\tau_1))Q(t-\tau_1) \\ \frac{dW}{dt} = -\gamma_2 W(t) + Ak(W(t-\tau_2))Q(t-\tau_2) \end{cases}$$

$$Q(t) = Q_0(t), W(t) = W_0(t), t \in [-\tau^*, 0], \tau^* = \max\{\tau_1, \tau_2\}$$

Delay  $\tau_1 \geq 0$  corresponds to the cell cycle duration.

Delay  $\tau_2 \geq 0$  corresponds to the amplification phase duration.

$$Q(t) \geq 0, W(t) \geq 0$$

Existence of nontrivial positive steady-state is ensured by:

$$(2^{-\gamma_1\tau_1} - 1)\beta(0) > k(0) + K \text{ and}$$

the function  $Q \mapsto Q\beta(Q)$  is decreasing in  $(Q_0, Q_1)$ , where

$$Q_0 = \beta^{-1} \left( \frac{k(0) + K}{2^{-\gamma_1\tau_1} - 1} \right) \text{ and } Q_1 = \beta^{-1} \left( \frac{K}{2^{-\gamma_1\tau_1} - 1} \right)$$

# Solution methods

# Solution methods

XPPAUT is "A tool for simulating, animating and analyzing dynamical systems." (G. B. Ermentrout)

B. Ermentrout, *Simulating, analyzing and animating dynamical systems: a guide to XPPAUT for researchers and students*, SIAM, 2002

<http://www.math.pitt.edu/~bard/xpp/xpp.html>

XPPAUT implementation of the methods:

	Expl.	Impl.	FS	AS	Stiff
Runge Kutta (RK)	+		+		
Dormand-Prince 5 (DP5)	+			+	
Rosenbrock (RB2)		+		+	+

*Rosenbrock* is based on Matlab version of the two step Rosenbrock algorithms.

*Delay equations* are solved by storing previous data and using cubic polynomial interpolation to obtain the delayed value.

*E. Hairer, (S.P. Norsett), G. Wanner, Solving ordinary differential equations I, II, Springer Ser. in Comp. Math., Springer, 2000 (part I), 2002 (part II)*

Motivation

Two leukopoiesis models

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# Clinical data

# Provided clinical data

Motivation

Two leukopoiesis models

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Concluding remarks

- Gathered amount of HSC (CD34+) – initial value for Q; Minimal required amount  $2 \times 10^6$  cells/kg, optimal  $5 \times 10^6$  cells/kg;
- After BMT – no blood system, i.e. initial values for matured cells are equal to 0;
- G-CSF is applied every day during the first month (NEUPOGEN – Filgrastim; GRANOCYTE – Lenograstim);
- Statistical data for T, B and NK cells and their subpopulations at several stages: before BMT (D), 1, 2, 3, 6, 9, 12, 18, 24 months after BMT.
- Diseases – Multiple Myeloma (MM), Non-Hodgkin's Lymphoma (NHL), Acute Myelogeneous Leukemia (AML)

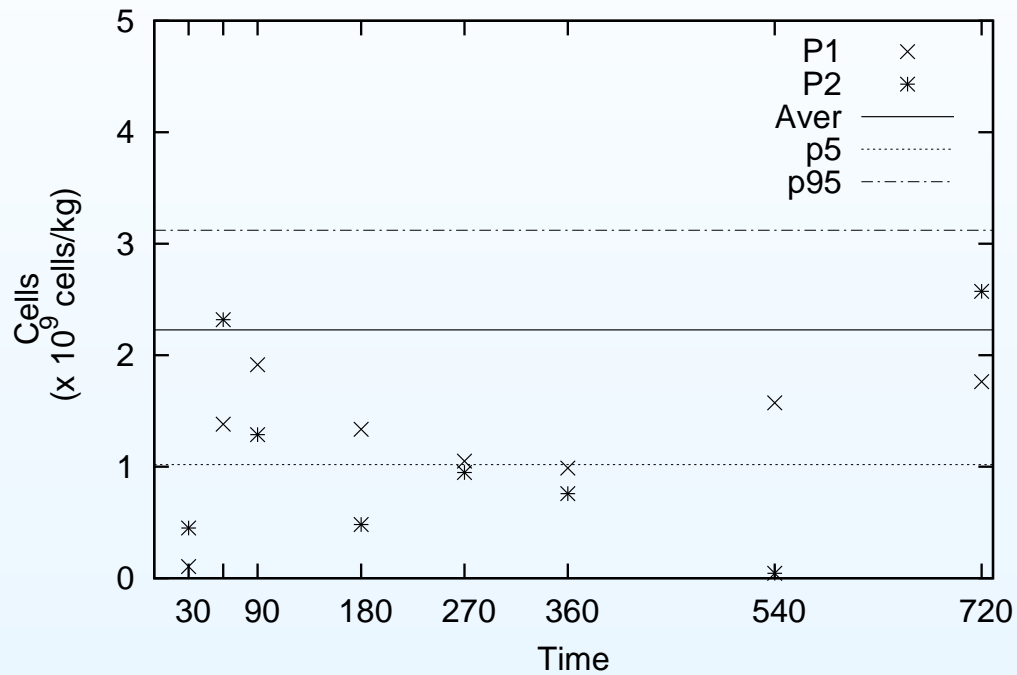
## 2 patients with AML

N	Date	Stage	Lymphocytes CD 45 AC	B cells CD19 AC	T cells CD3 AC	NK cells CD 56 AC	$T_n$ AC
P1	11/11/05	D	571	5	491	33	114.36
	01/12/05	E1	104	1	90	8	27.8
	16/01/06	E2	1382	15	1172	104	222.83
	06/02/06	E3	1914	57	1501	226	249.66
	10/05/06	E6	1336	123	1017	107	232.02
	30/08/06	E9	1049	156	703	103	75.7
	14/11/06	E12	988	270	600	121	108.01
	09/05/07	E18	1573	267	959	215	215.84
	21/10/08	E 24	1763	924	924	400	333
P2	11/10/06	D	1946	25	1721	53	160.01
	01/11/06	E1	450	30	410	19	39.7
	07/12/06	E2	2319	32	2082	121	103.35
	17/01/07	E3	1287	97	1061	84	92.55
	25/04/07	E6	481	62	380	25	20.19
	04/07/07	E9	947	9	886	31	155.05
	17/10/07	E12	759	1	729	17	115.22
	27/05/08	E 18	44	0	43	1	23.41
	16/10/08	E 24	2572	2279	2291	208	182.61



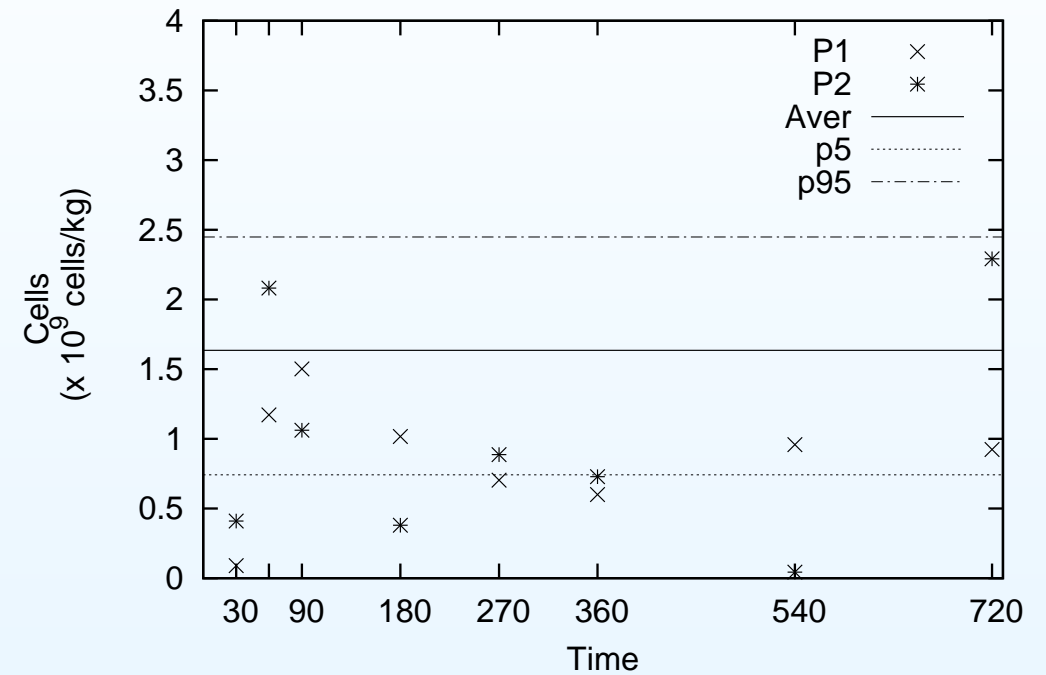
# Healthy controls vs patients with AML

Healthy controls /AML - Lymphocytes



Lymphocytes (CD 45 AC)

Healthy controls /AML - T cells



T cells (CD 3 AC)

# Numerical tests

# Model parameters

## GFM

$$\beta(E) = \beta_0 \frac{E}{1 + E}, \quad \beta_0 > 0$$

$$g(Q) = GQ, \quad G > 0$$

$$f(M) = \frac{a}{1 + KM^r}, \quad a, K > 0, r > 0$$

Param	Value	Range ( $day^{-1}$ )
$\delta$	0.01 $day^{-1}$	0 – 0.09
$G$	0.04 $day^{-1}$	0 – 0.09
$\beta_0$	0.5 $day^{-1}$	0.08 – 2.24
$\gamma$	0.2 $day^{-1}$	0 – 0.9
$\mu$	0.02 $day^{-1}$	0.001 – 0.1
$k$	2.8 $day^{-1}$	—
$a$	6570	—
$K$	0.0382	—
$r$	7	—

## LM

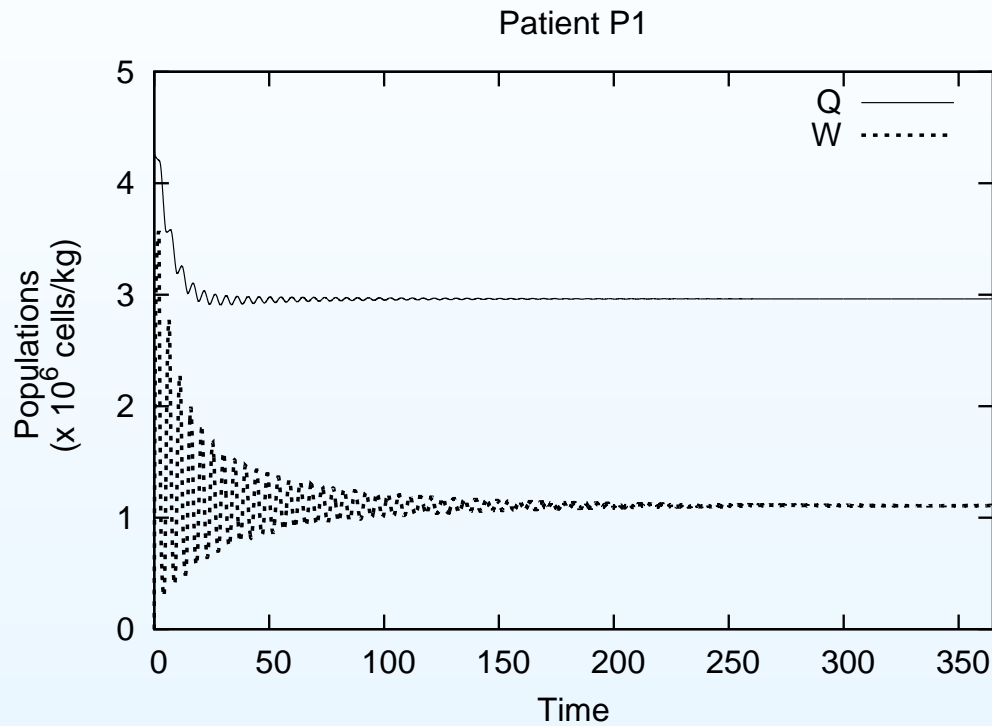
$$\beta(Q) = \frac{\beta_0}{1 + Q^n}, \quad \beta_0 > 0$$

$$k(W) = \frac{k_0}{1 + W^m}, \quad k_0 > 0$$

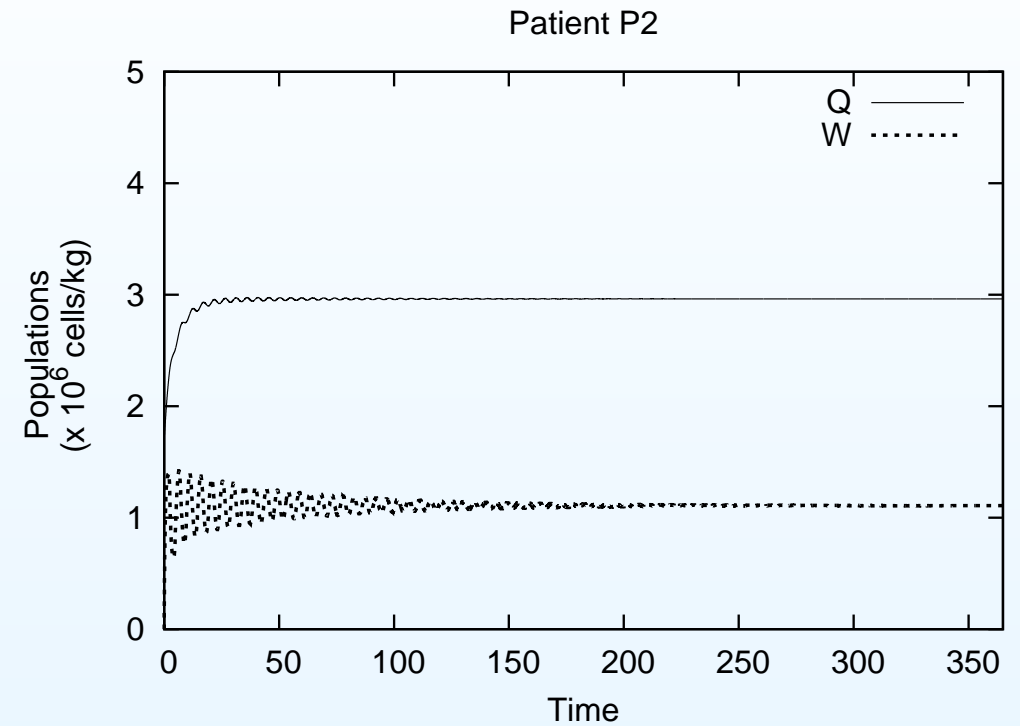
$$A = \alpha 2^i, \quad \alpha \in (0, 1)$$

Param	Value
$\beta_0$	1.77 $day^{-1}$
$k_0$	0.1 $day^{-1}$
$n$	3
$m$	2
$\gamma_1$	0.1 $day^{-1}$
$\gamma_2$	2.4 $day^{-1}$
$K$	0.02 $day^{-1}$
$A$	20

# Results $Q(t)$ and $W(t)$ , model data from paper LM

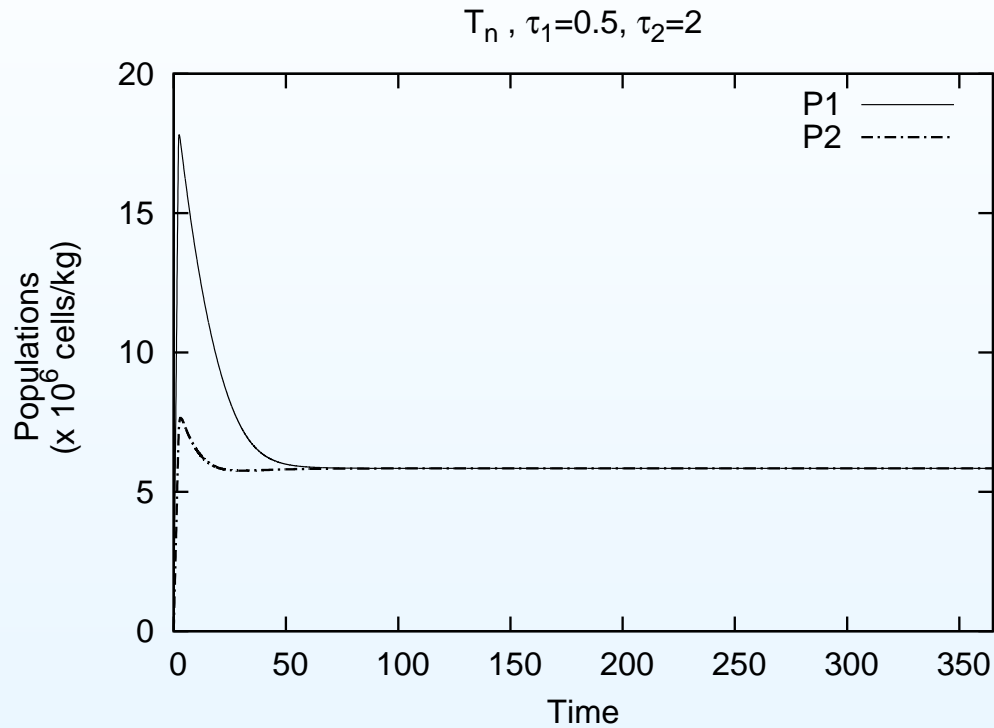


Patient P1:  
 $Q(0) = 4.32(\times 10^6)$  cells/kg,  
 $W(0) = 0$

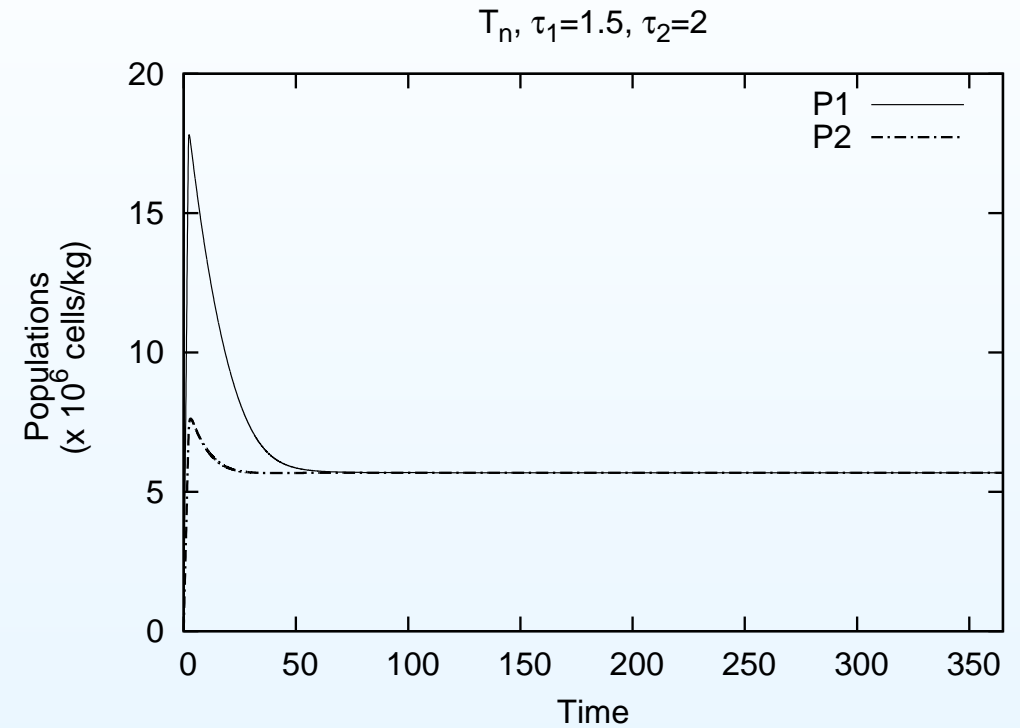


Patient P2:  
 $Q(0) = 1.69(\times 10^6)$  cells/kg,  
 $W(0) = 0$

# Results $W(t)$ , LM – varying $\tau$



$$\tau_1 = 0.5, \tau_2 = 2.$$

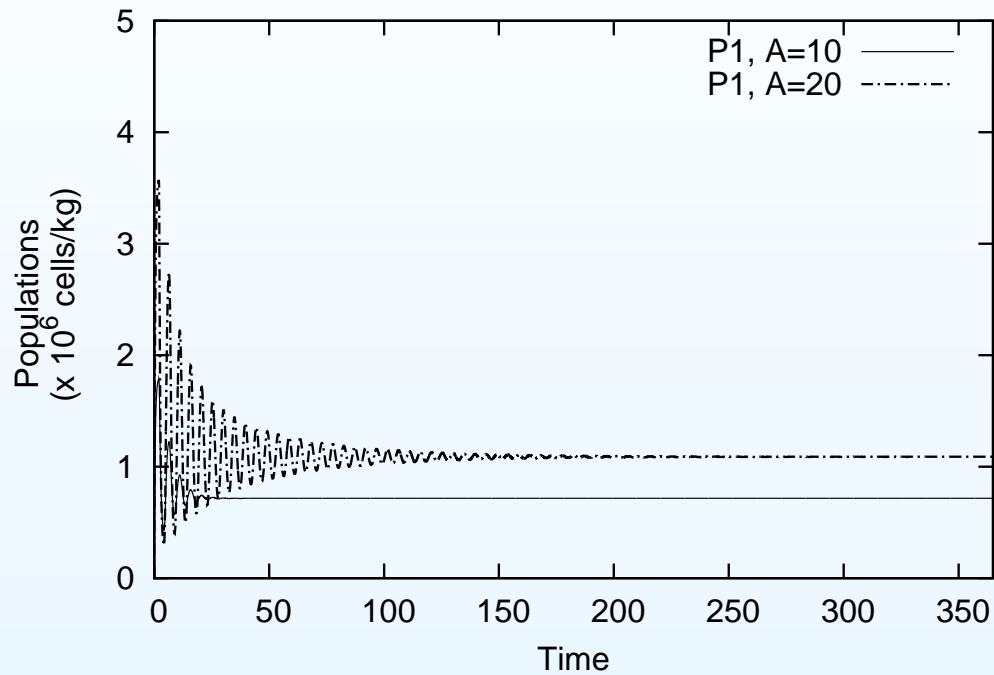


$$\tau_1 = 1.5, \tau_2 = 2.$$

P1 and P2 –  $T_n$  cells with  $\gamma_2 = 0.04$  (Moore, Li (2004))

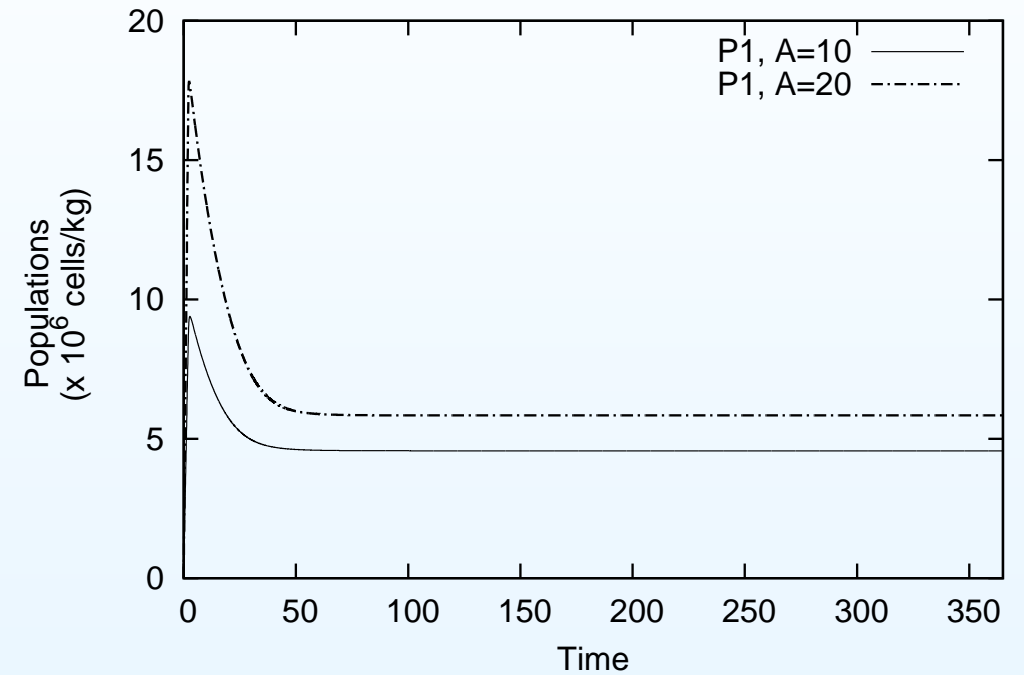
# Results, Patient 1, LM – varying $A$

W,  $\tau_1=0.5, \tau_2=2$



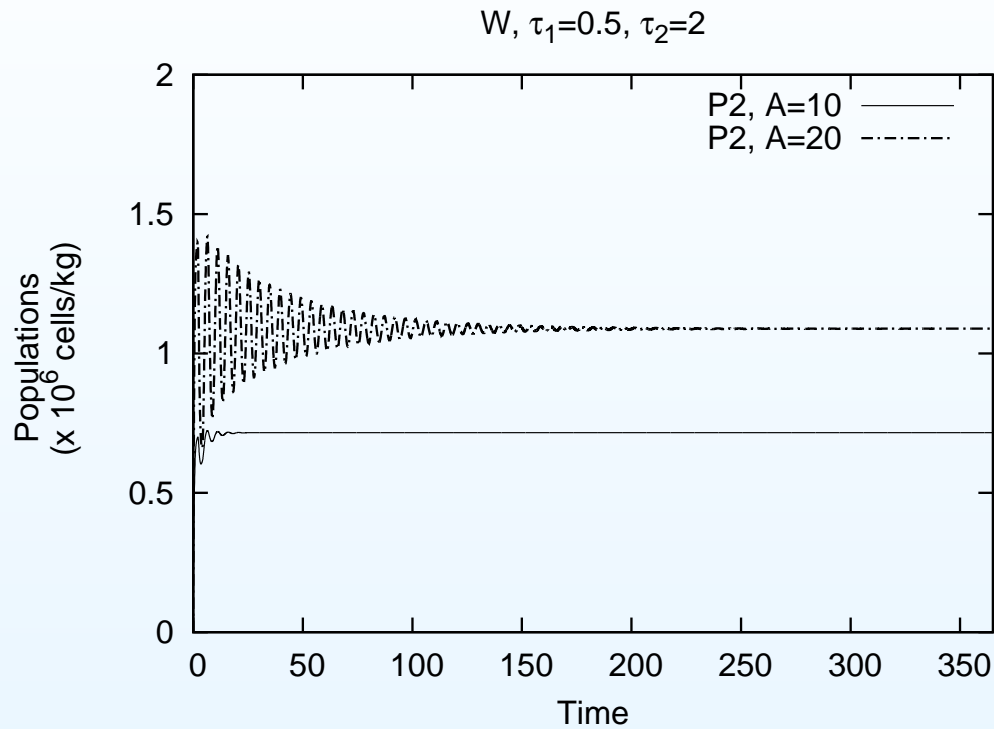
$\gamma_2 = 2.4, A = 10, 20$

T<sub>n</sub>,  $\tau_1=0.5, \tau_2=2$

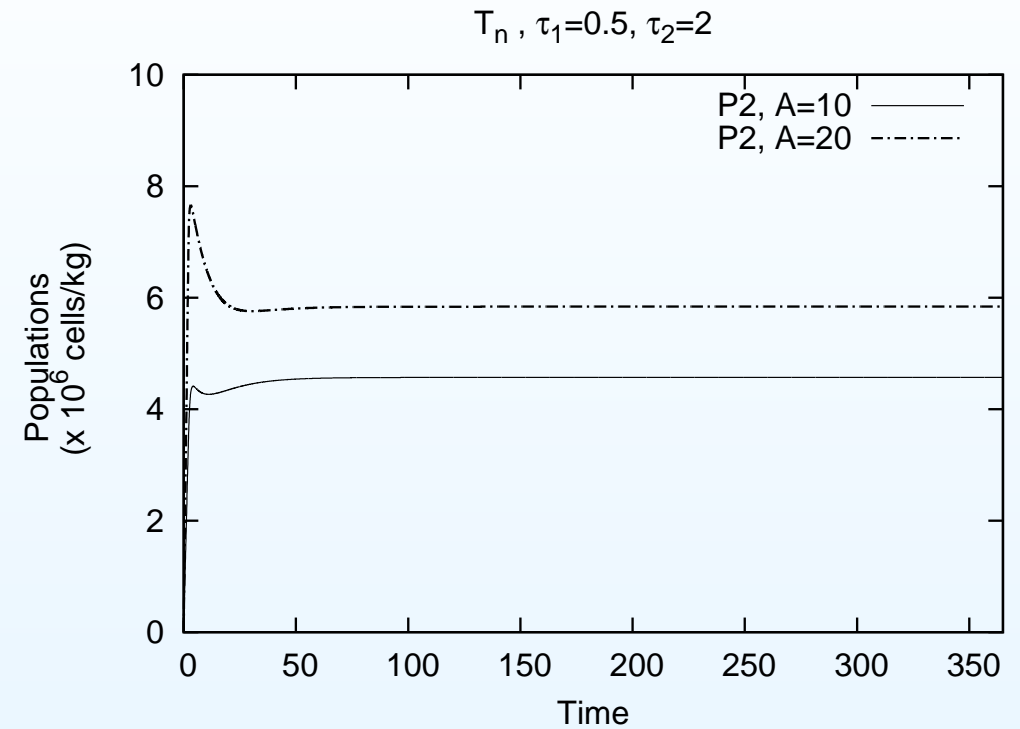


$\gamma_2 = 0.04, A = 10, 20$

# Results, Patient 2, LM – varying $A$



$$\gamma_2 = 2.4, A = 10, 20$$



$$\gamma_2 = 0.04, A = 10, 20$$

## Concluding remarks



# Concluding remarks

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Concluding remarks

- Open issues
  - Why does the GFM model "crash" with the clinical data?
  - Which parameters/functions should be changed and how in order to have steady states of LM closer to the clinical data?
- Further steps – sensitivity analysis and parameter estimation of both models

*Clinical data is provided by Dr. M. Guenova and Dr. L. Gartcheva from NCHT*

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Thank you for your attention!